

## ORIGINAL ARTICLE

# Herbicide resistance-endowing ACCase gene mutations in hexaploid wild oat (*Avena fatua*): insights into resistance evolution in a hexaploid species

Q Yu<sup>1</sup>, MS Ahmad-Hamdani<sup>1,2</sup>, H Han<sup>1</sup>, MJ Christoffers<sup>3</sup> and SB Powles<sup>1</sup>

Many herbicide-resistant weed species are polyploids, but far too little about the evolution of resistance mutations in polyploids is understood. Hexaploid wild oat (*Avena fatua*) is a global crop weed and many populations have evolved herbicide resistance. We studied plastidic acetyl-coenzyme A carboxylase (ACCase)-inhibiting herbicide resistance in hexaploid wild oat and revealed that resistant individuals can express one, two or three different plastidic ACCase gene resistance mutations (Ile-1781-Leu, Asp-2078-Gly and Cys-2088-Arg). Using ACCase resistance mutations as molecular markers, combined with genetic, molecular and biochemical approaches, we found in individual resistant wild-oat plants that (1) up to three unlinked ACCase gene loci assort independently following Mendelian laws for disomic inheritance, (2) all three of these homoeologous ACCase genes were transcribed, with each able to carry its own mutation and (3) in a hexaploid background, each individual ACCase resistance mutation confers relatively low-level herbicide resistance, in contrast to high-level resistance conferred by the same mutations in unrelated diploid weed species of the Poaceae (grass) family. Low resistance conferred by individual ACCase resistance mutations is likely due to a dilution effect by susceptible ACCase expressed by homoeologs in hexaploid wild oat and/or differential expression of homoeologous ACCase gene copies. Thus, polyploidy in hexaploid wild oat may slow resistance evolution. Evidence of coexisting non-target-site resistance mechanisms among wild-oat populations was also revealed. In all, these results demonstrate that herbicide resistance and its evolution can be more complex in hexaploid wild oat than in unrelated diploid grass weeds. Our data provide a starting point for the daunting task of understanding resistance evolution in polyploids. *Heredity* advance online publication, 10 October 2012; doi:10.1038/hdy.2012.69

**Keywords:** ACCase mutation; *Acc1*; herbicide resistance; hexaploid wild oat (*Avena fatua*); polyploidy; resistance evolution

## INTRODUCTION

About 30–70% of plant species are of polyploid origin, and many herbicide-resistant weed species are polyploids. However, our understanding on the evolution of herbicide resistance in polyploid weeds is rather limited. Wild oats (*A. fatua* and *A. sterilis*) are self-pollinated allohexaploid species of the Poaceae (grass) family and are among the world's most common and economically damaging weeds in crop production. Acetyl-coenzyme A carboxylase (ACCase)-inhibiting herbicides have been widely used for wild oat control. ACCase (EC 6.4.1.2) is a key enzyme in most organisms, catalyzing the first step of fatty acid biosynthesis. Herbicidal inhibition of plant ACCase is also important in world agriculture for weed control. Three chemically distinct classes of herbicides that inhibit ACCase are the aryloxyphenoxypropionates, the cyclohexanediones and the more recent phenylpyrazolin class consisting of pinoxaden herbicide. These three chemical classes are hereafter collectively referred to as ACCase herbicides. In plants, ACCase in plastids is essential for the biosynthesis of primary fatty acids and cytosolic ACCase is involved in biosynthesis of long-chain fatty acids (Shorosh *et al.*, 1994). Homomeric ACCase in the cytosol of nearly all plant species, and

heteromeric ACCase in the chloroplasts of dicots, are relatively insensitive to ACCase herbicides. In contrast, the plastidic homomeric ACCase found in nearly all grass species is ACCase herbicide sensitive, and thus grass species are generally controlled by ACCase herbicides.

Global and often intensive ACCase herbicide use has selected for resistance in many grass weeds (Heap, 2011), since the first report by Heap and Knight (1982). Biochemical mechanisms known to endow ACCase herbicide resistance in plants are target-site ACCase gene mutations and/or non-target-site enhanced rates of ACCase herbicide metabolism (reviewed by Devine and Shimabukuro, 1994; Délye, 2005; Powles and Yu, 2010). Thus far, among grass weeds, nine distinct amino-acid substitutions in the carboxyl transferase domain of plastidic ACCase genes have been identified as individually endowing target-site resistance to certain ACCase herbicides (reviewed by Délye *et al.*, 2005; Powles and Yu, 2010; see also Collavo *et al.*, 2011). In diploid grass species, these individual ACCase gene resistance mutations confer moderate to high-level resistance to ACCase herbicides (Délye *et al.*, 2005; Yu *et al.*, 2007). Depending on specific ACCase mutations, herbicides and herbicide rates, ACCase resistance alleles in diploid grass species have been found to be

<sup>1</sup>Australian Herbicide Resistance Initiative, School of Plant Biology, Faculty of Natural and Agriculture Sciences, University of Western Australia, Crawley, Western Australia, Australia; <sup>2</sup>Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia, Serdang, Malaysia and <sup>3</sup>Department of Plant Sciences, North Dakota State University, Fargo, ND, USA.

Correspondence: Professor S Powles, Australian Herbicide Resistance Initiative, School of Plant Biology, Faculty of Natural and Agriculture Sciences, University of Western Australia, Crawley, Western Australia 6009, Australia.

E-mail: stephen.powles@uwa.edu.au

Received 4 November 2011; revised 27 July 2012; accepted 1 August 2012

partially dominant or dominant over wild-type alleles (Christoffers *et al.*, 2002; Tal and Rubin, 2004; Yu *et al.*, 2007; Scarabel *et al.*, 2011).

Many resistant wild-oat populations have been reported in at least 12 countries (Heap, 2011). Early studies of the mechanism(s) endowing resistance to ACCase herbicides in resistant *Avena* spp. populations revealed resistant ACCase (Maneechote *et al.*, 1994, 1997; Seefeldt *et al.*, 1996; Shukla *et al.*, 1997; Cocker *et al.*, 2000) with more recent reports identifying specific ACCase gene mutations conferring the resistance (Christoffers *et al.*, 2002; Liu *et al.*, 2007; Cruz-Hipolito *et al.*, 2011; Beckie *et al.*, 2012). However, hexaploidy in *Avena* and other polyploid species renders identifying and understanding herbicide resistance difficult. For herbicide resistance, these previous studies investigating the biochemical and molecular basis of ACCase herbicide resistance in *Avena* spp. did not investigate the complexities of gene transcription/expression in a polyploid species. Whereas in diploids both gene copies are usually transcribed and expressed, expression may be far more complicated in polyploid species. Hexaploid wild oat ( $2n=6\times=42$ ) has a genome structure of AACDD originating through allopolyploidy involving three diploid species (Rajhathy and Morrison, 1960; Jellen *et al.*, 1994; Yang *et al.*, 1999). *A. longiglumis* has been suggested as a possible A-genome donor in the evolution of hexaploid *Avena* spp. (Nikoloudakis *et al.*, 2008), while the likely C-genome donor is *A. ventricosa* (Nikoloudakis and Katsiotis, 2008). The origin of the D genome is less clear, but it is more similar to the A genome than the C genome (Yang *et al.*, 1999). All hexaploid *Avena* spp. show bivalent chromosome pairing with disomic inheritance (Thomas, 1992). Therefore, each gene is predicted to have three homoeologous copies, one from each of the three original genomes. While there is increasing understanding of homoeolog genetics and expression/silencing in allotetraploid *Arabidopsis* and polyploid crop species such as *Brassica* and wheat (reviewed by Wendel, 2000; Adams, 2007; Chen, 2007), all that is currently known for *A. fatua* plastidic ACCase genes is that there are three gene copies (*Acc1;1*, *Acc1;2* and *Acc1;3*, Christoffers *et al.*, 2002), and it is rarely known whether all homoeologous genes are expressed or how they interact and are inherited. In studying the molecular and biochemical basis of herbicide resistance in four hexaploid wild-oat (mainly *A. fatua*) populations, we identified individuals with one, two or three distinct resistance-endowing ACCase gene mutations. These multiple-mutation individuals provided a valuable opportunity to study gene expression and inheritance relating to natural selection in polyploids. Using resistance gene mutations as markers, we investigated homoeologous ACCase gene inheritance, transcription and *in vitro* enzyme activity in hexaploid *Avena*. Using herbicide resistance evolution as an example, we provide insights into resistance evolution in hexaploid wild oat.

## MATERIALS AND METHODS

### Plant material

Herbicide-resistant wild-oat populations M3/4, M3/5, M3/27 (*A. fatua*) and M5W/7 (*A. sterilis*), hereinafter called R1, R2, R3 and R4, respectively (Ahmad-Hamdani *et al.*, 2011), were used in this study along with a susceptible *A. fatua* population (Owen and Powles, 2009). Plants surviving high rates of commercial ACCase herbicides diclofop-methyl fenoxaprop-p-ethyl, or sethoxydim, (all from Bayer CropScience, Melbourne, VIC, Australia) from each population enabled identification of specific ACCase resistance mutations among individual plants. These individuals with a specific ACCase mutation were grown to maturity in a glasshouse and allowed to self-pollinate and produce seeds. To ensure no chance of cross-pollination, individual plants were insulated in micro-perforated plastic bags and seeds from single plants were harvested separately. These single-plant-derived progeny lines with known ACCase resistance mutation(s) were used for subsequent studies.

### Plant growth and herbicide treatment

Scarified seeds of the resistant progeny lines were germinated on agar (0.6% w/v) and seedlings transplanted into potting mix and grown in a controlled environment room with a day/night temperature of 20/15 °C and a 12-h photoperiod (500 μmol photons m<sup>-2</sup> s<sup>-1</sup>). When seedlings reached the 3- to 4-leaf stage, one leaf from each plant was sampled for ACCase sequencing and mutation genotyping before the plants were herbicide treated for resistance phenotyping. The susceptible population was treated in the same manner. Individuals of the R progeny lines specifically identified to possess a known resistance-endowing single-ACCase mutation (Ile-1781-Leu, Asp-2078-Gly or Cys-2088-Arg) and the susceptible population were treated with a commercial formulation of the ACCase herbicide diclofop-methyl (delivered in 112 l ha<sup>-1</sup> water at a pressure of 200 kPa). Diclofop-methyl was used at 2000 g ha<sup>-1</sup>, which fully controlled the S population. Plants were returned to the controlled environment room after treatment and visual assessment for resistance and susceptibility was made 3 weeks after treatment. Plants were recorded as resistant if they displayed strong growth after herbicide treatment and as susceptible if they displayed severe symptoms of leaf chlorosis, desiccation, retarded growth or plant death, similar to the S population.

### ACCase gene sequencing

Genomic DNA (gDNA) was extracted from the leaf tissues of resistant and susceptible plants according to Yu *et al.* (2008) and the regions in the wild-oat plastidic ACCase carboxyl transferase domain with known and potential ACCase resistance mutation sites were amplified using published primer pairs SQCT-β1 F/SQCT-β1 R and SQCT-α1 F/SQCT-α1 R for wild oat (Liu *et al.*, 2007 and Table 1) under modified PCR conditions. The PCR was conducted in a 25-μl volume that consisted of 1–2 μl of gDNA, 0.5 μM of each primer, and 12.5 μl of 2 × GoTaq Green Master Mix (Promega, Madison, WI, USA). The PCR was run with the following profile: 94 °C for 4 min; 35–40 cycles of 94 °C for 30 s, X °C for 30 s, and 72 °C for 30–120 s; followed by a final extension step of 5–7 min at 72 °C. X refers to the respective annealing temperature used for each primer pair (Table 1). The PCR products were purified from agarose gels using the Wizard SV Gel and PCR Clean-up System (Promega), and sequenced using a commercial sequencing service. All sequence chromatograms were visually checked for quality and consistency before sequences were assembled and aligned.

### ACCase resistance mutation genotyping and segregation analysis using PCR-based, non-homoeolog-specific molecular markers

Published derived cleaved amplified polymorphic sequence (dCAPS) markers for mutations at positions 1781 (Kaundun and Windass, 2006) and 2078 (Kaundun, 2010) were used in this study with modified primers (Table 1) and PCR conditions as described above. In addition, based on the ACCase sequence information obtained from the bulked susceptible (20 plants) and a number of individual R samples (at least 35), we developed a CAPS marker for the 2088 mutation (Table 2). The nucleotide T to C mutation at codon 2088, causing a Cys to Arg amino-acid substitution, creates a *Hae*II restriction site (Table 2). Sequence results revealed no other single-nucleotide polymorphisms around this restriction site. Therefore, the primer pair SQCT-α1F and SQCT-α1R (Table 1) was used to amplify a 602-bp fragment, which was then digested with *Hae*II. Wild-type 2088-Cys alleles would generate a single undigested 602 bp band, while mutant resistant 2088-Arg alleles would generate a 529 bp band, plus an undetectable 73 bp fragment. Heterozygous ACCase loci would produce both the 602 and 529 bp bands. Restriction digestions were carried out according to the manufacturer's recommendations (New England BioLabs, Ipswich, MA, USA) and digestion patterns were viewed on 2% agarose gels (electrophoresis at 90–100 V for 120–140 min) stained with ethidium bromide. These three (d)CAPS markers, for codons 1781, 2078 and 2088, were used to analyze segregation patterns for ACCase resistance mutation(s) in the R2-resistant progeny lines. While these (d)CAPS markers are codominant with wild-type alleles, they do not discriminate among ACCase gene copies and, therefore, were considered dominant markers (unable to detect true allelic heterozygosity) for the purposes of analyzing plant genotypes and segregation patterns (see discussions of homoeoallelic heterozygosity below).

**Table 1** Primer pairs (resistance mutation site numbering corresponds to amino acid sequence of full-length plastidic ACCase in *A. myosuroides*)

Primer pair	Sequence (5'-3')	Annealing temperature used (°C)	Usage	Product size (bp)	Targeted mutation site
SQCT-β1F	CATCATTTTCTGTATGCCAGTGGG	65	Sequencing	999	-1781-
SQCT-β1R	CTGTATGCACCGTATGCCAAG				
SQCT-α1F	AATACATGTGATCCTCGTGACAG	60	Sequencing	602	-1999–2096-
SQCT-α1R	TCCTCTGACCTGAACCTTGATCTC				
Nsil1781F	CTGAATGAAGAAGACTATGGTCG	60	dCAPS	165	1781
Nsil1781R	AGAATACGCACTGGCAATAGCAGCACTTCCATGCA				
Rsal2078F	TTCTCTGGTGGGCAAAGAGACCTTTTGAAGG	62	dCAPS	181	2078
Rsal2078R	CATAGCACTCAATGCGATCTGGATTATCTTGTA				
SQCT-α1F	See above	60	CAPS	602	2088
SQCT-α1R	See above				
Nsil1781F	See above	60	cDNA cloning	1122	-1781–2096-
SQCT-α1R	TCCTCTGACCTGAACCTTGATCTC				
Nsil1781F	See above	60	cDNA sequencing	1050	-1781–2096-

Abbreviations: ACCase, acetyl-coenzyme A carboxylase; cDNA, complementary DNA; dCAPS, derived cleaved amplified polymorphic sequence.

**Table 2** (d)CAPS markers used for identification of ACCCase mutant alleles in wild-oat populations. Ambiguity nucleotide codes: R = A/G, Y = C/T

Target mutation	Primer pair used	Type	Restriction enzyme	Recognition site	Fragment size (bp)	
					Susceptible (S)	Resistant (R)
1781	Nsil1781F/ Nsil1781R	dCAPS	<i>Nsil</i>	5'-ATGCA^T-3'	130, 35	165
2078	Rsal2078F/ Rsal2078R	dCAPS	<i>Rsal</i>	5'-GT^AC-3'	181	147, 34
2088	SQCT-α1 F/ SQCT-α1 R	CAPS	<i>HaeIII</i>	5'-RGC^G^Y-3'	602	529,73

Abbreviations: ACCCase, acetyl-coenzyme A carboxylase; dCAPS, derived cleaved amplified polymorphic sequence.

### ACCCase complementary DNA (cDNA) fragment cloning and sequencing

Total RNA was extracted from leaf tissue of individual plants possessing all three ACCCase mutations (1781 + 2078 + 2088) using the Plant RNeasy Mini Kit (Qiagen, Hilden, Germany). gDNA contamination was removed using the TURBO-DNA free kit (Ambion, Austin, TX, USA), which is highly effective in removing trace quantities of DNA contamination. Oligo(dT)-primed first-strand cDNA was prepared from 2 µg total RNA using the Omniscript Reverse Transcription system (Qiagen). The primers Nsil1781F (modified from Kaundun and Windass, 2006) and SQCT-α1R were paired to amplify a 1122 bp cDNA fragment containing all known resistance-endowing ACCCase gene mutation sites (1781, 1999, 2027, 2041, 2078, 2088 and 2096), as well as polymorphic sites for the three ACCCase gene copies in *A. fatua* (Christoffers *et al.*, 2002). This fragment was cloned into the pGEM-T vector (Promega) and transformed into *E. coli*. White colonies with putative inserts were used as templates for PCR re-amplification of the 1122 bp fragment and screened for the 1781, 2078 or 2088 mutations using PCR-based markers (see below). Fragments that contained putative 1781, 2078 or 2088 mutations were selected for confirmatory sequencing using two forward primers, SQCT-α1F and Nsil1781F (Table 1), with overlapping amplification regions.

### In vitro inhibition of ACCCase activity by ACCCase herbicides

The leaf tissue from R2 progeny lines determined to be homozygous (no dCAPS/CAPS marker segregation) for one or two ACCCase mutation(s), or from plants of segregating progeny lines heterozygous for two or three mutations were used for *in vitro* ACCCase activity assays. Leaf material at the

3 to 4-leaf seedling stage (unless otherwise specified) was harvested, snap frozen in liquid nitrogen and stored at -80 °C until use. ACCCase extraction, partial purification and *in vitro* enzyme inhibition by ACCCase herbicides was performed according to Yu *et al.* (2004). Two subsamples from each extraction were assayed, and there were at least two extractions per population per herbicide treatment. Technical grade diclofop acid, fenoxaprop acid and clethodim (94–98% purity, Bayer CropScience, and Sumitomo Chemical Australia, Epping, NSW, Australia) and sethoxydim (52%, Bayer CropScience) were used in assays to determine the herbicide concentration causing 50% inhibition of ACCCase activity ( $I_{50}$ ). Herbicide stock solutions were prepared in 100 mM Tricine buffer. Stock solutions of diclofop or fenoxaprop contained 10% acetone with final concentrations of <1% acetone in each assay mix. Tissue protein content was determined by the Bradford method (Bradford, 1976).

The *in vitro* ACCCase assay in *A. fatua* was reported to be affected by experimental conditions such as the composition of the extraction buffer and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> precipitation of the protein (Shukla *et al.*, 1997). In order to test if different experimental conditions affected our resistant: susceptible  $I_{50}$  ratios, we determined  $I_{50}$  values when (1) ACCCase was assayed with and without desalting, (2) when crude protein extract was precipitated with 10–40% versus 40% ammonium sulfate saturation and (3) when a different extraction buffer (buffer D in Shukla *et al.*, 1997) was used. The herbicide  $I_{50}$  values were found to vary in each case for resistant and susceptible samples, but the  $I_{50}$  resistant/ susceptible ratios (referred to as R/S ratios) remained similar. In addition, plants from purified *L. rigidum* mutant lines homozygous for ACCCase mutations 1781, 2078 or 2088 (Yu *et al.*, 2007) were included as positive controls for the assay.

### Statistical analysis

The  $I_{50}$  was estimated by non-linear regression using the logistic model:

$$y = C + \frac{D - C}{1 + (x/ED_{50})^b}$$

where  $C$  = lower limit,  $D$  = upper limit,  $ED_{50}$  = dose giving 50% response and  $b$  = slope around  $ED_{50}$ . Estimates were obtained using Sigmaplot (version 12.0, Systat Software Inc., San Jose, CA, USA). Data was subjected to one-way analysis of variance and significant differences in ACCase activities and  $I_{50}$  values among genotypes were determined by Duncan's test using SPSS software (version 11.5, SPSS Inc., Chicago, IL, USA). Data sets from different experiments were analyzed separately if significant differences were found because of differences in experimental set up or plant developmental stages.

Independent assortment of mutations was evaluated using logarithm of the odds score analysis to detect genetic linkage (Strachan and Read, 1999). This analysis compares the likelihood of two loci being linked versus unlinked (independent assortment). An expected Mendelian disomic inheritance segregation ratio of 27:9:9:3:3:3:1 was predicted from the self-pollination of a parent heterozygous for three different and unlinked dominant (d)CAPS markers. Likewise to test for polysomic inheritance, an expected segregation ratio of 147:69:69:69:15:15:15:1 was predicted for self-pollination of a hexaploid parent carrying three different (d)CAPS markers and three wild-type loci, where all six gene loci are unlinked and segregate randomly. In the polysomic model, the (d)CAPS markers were codominant among themselves but dominant over wild-type loci, and no chromatid segregation was considered (no double reduction).

Owing to relatively small sample sizes in dCAPS/CAPS marker segregation experiments, the exact test for goodness of fit for disomic and polysomic inheritance models was performed using the SAS software (version 9.3 for Windows, SAS Institute Inc., Cary, NC, USA) (program according to online Handbook of Biological Statistics) to analyze segregation ratios of different molecular genotypes in the R2 progeny lines.

## RESULTS

### ACCase gene sequencing revealed that individual plants can possess from one to three different ACCase resistance mutations

At least 38 individual R plants from each of the four parent R populations were sequenced, resulting in the identification of three known ACCase gene resistance-endowing mutations; Ile-1781-Leu, Asp-2078-Gly and Cys-2088-Arg (Table 3). No other known ACCase resistance mutations were found. The 2078 mutation was the most common as it was found in resistant individuals within all four populations. The 2088 mutation was present only in individuals of two resistant populations (R2 and R3). The 1781 mutation was found only in one population (R2). Importantly in this R2 population, many individuals displayed more than one ACCase gene mutation, with 32% of individuals possessing two mutations, and one plant with all three resistance mutations (Table 3). While the majority of the

plants in populations R2 and R3 had ACCase mutations, 89% of the resistant plants in population R1, and 26% in population R4, did not contain any of the known ACCase gene mutations (Table 3). These plants lacking ACCase mutations were found to possess non-target-site based resistance mechanisms (enhanced rates of herbicide metabolism) (Ahmad-Hamdani *et al.*, 2012) unrelated to ACCase genes.

### (d)CAPS marker analysis revealed homoeoallelic heterozygosity for ACCase resistance mutations in individual parent resistant plants

ACCase gene sequencing showed that individual parent resistant plants possessing resistance-endowing mutation(s) always displayed double peaks, that is, both mutant and wild-type nucleotides, at mutation sites in the sequence chromatograms (data not shown). Similarly, when these sequenced individuals were analyzed using the three (d)CAPS markers, plants either displayed both resistant mutant and wild-type bands (RS) or only susceptible wild-type bands (S) (Figure 1). This is in contrast to the genotype patterns (R, RS and S) usually detected in diploid species by the same markers (Kaundun and Windass, 2006; Kaundun, 2010). The most likely reason for consistent presence of wild-type nucleotides/bands at resistance mutation sites is the amplification of more than one homoeologous ACCase genes by the primers used in our assays, that is, homoeologous heterozygosity. Sequencing and (d)CAPS marker analysis would be expected to always reveal wild-type nucleotides/bands unless the same mutation was found in a homozygous state among all homoeologs. Homoeologous heterozygotes may or may not have allelic heterozygosity. We consider fixed heterozygosity, where there is polymorphism among homoeologs but each homoeolog itself is homozygous (no allelic heterozygosity), to be a specific type of homoeologous heterozygosity.

### (d)CAPS marker segregation analysis of progeny plants revealed three homoeologous ACCase gene loci, each segregating and assorting independently

To explore parental genotypes and segregation patterns in more detail, individual R2 population plants possessing one mutation (1781, 2078 or 2088), two mutations (1781 + 2078 or 2078 + 2088) or three mutations (1781 + 2078 + 2088) were selfed and seeds from single plants harvested for subsequent study. If the parent plants identified by marker analysis were true allelic heterozygotes for a single resistance mutation, that is, having both wild-type and mutant alleles at the same locus (homoeologous copies are considered different loci); and the two alleles segregated at a single ACCase gene locus, then heterozygous RS (allelic and homoeologous) and homozygous S

**Table 3** ACCase carboxyl transferase domain sequencing results showing one, two or three different mutations in resistant individual plants of four wild-oat (*Avena* spp.) populations

Resistant Population	Number of plants analyzed	Number of individuals in which specific mutation(s) were identified							
		1781 (Ile-Leu)	2078 (Asp-Gly)	2088 (Cys-Arg)	1781 + 2078	1781 + 2088	2078 + 2088	1781 + 2078 + 2088	Nil
R1	38	0	4	0	0	0	0	0	34
R2	38	2	10	12	1	8	3	1	1
R3	38	0	34	1	0	0	0	0	3
R4	43	0	32	0	0	0	0	0	11

Abbreviation: ACCase, acetyl-coenzyme A carboxylase. Plants without identified mutations are suspected as having non-target-site resistance mechanism(s).

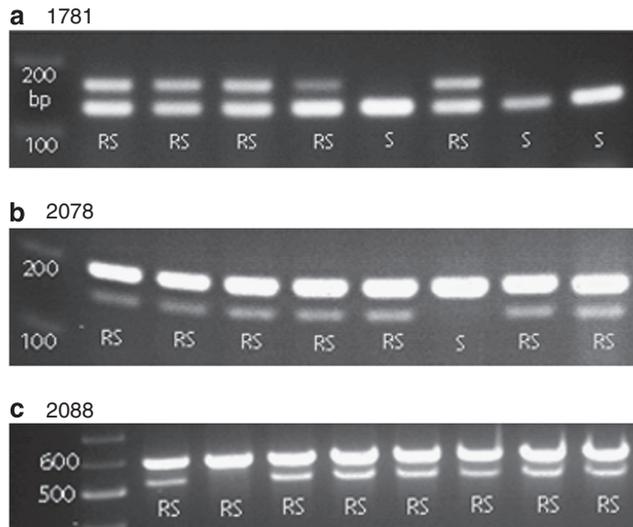
genotypes would be marker-detected among the progeny of RS-selfed plants. When this analysis was done for the single-mutation progeny lines carrying 1781, 2078 or 2088 mutation, only the RS (but no S) genotype was detected among the progeny (Table 4, Figure 2). As at least 18 ( $n \geq 18$ ) individual progeny from each line were analyzed, the probability of missing a homozygous S genotype among the progeny is low ( $P = 0.75^n = 0.006$ ). These results established that the single-mutation hexaploid *Avena* parent plants were homozygous for

their particular resistance mutation (that is, AA\_ \_ \_ \_ , BB\_ \_ \_ \_ or CC\_ \_ \_ \_ genotypes in Table 4, if A = 1781, B = 2078 and C = 2088). The fixed heterozygosity (RS) detected in these plants by the molecular markers is due to the susceptible allele(s) or wild-type nucleotide being amplified from other ACCase gene loci (homoeoallelic heterozygosity). The growth of all RS genotypes was unaffected by diclofop treatment ( $2000 \text{ g ha}^{-1}$ ), and no susceptible plants were found, establishing the RS genotypes with allelic R mutation homozygosity as diclofop resistant (Table 4).

For the selfed plants containing the two mutations 1781 + 2088, all progeny plants examined (24) possessed both the 1781 and 2088 mutation and all were scored RS due to homoeologous heterozygosity (Table 4). The lack of S-genotype progeny indicates no allelic segregation for either of the two mutations, again likely because the parent plants were homozygous for the two mutations at different ACCase gene loci (that is, AACC\_ \_ \_ in Table 4).

For the selfed plants containing the two mutations 2078 + 2088, 27 out of 32 progeny plants also had both mutations while five had only 2078 but not the 2088 mutation. This indicates that there was no allelic segregation for the 2078 mutation, but the plants did segregate for the 2088 mutation (as 2088 RS and 2088 S) (Table 4). The observed ratio of these 2088 segregates, RS:S = 27:5, was a good fit ( $P = 0.54$ ) to the Mendelian 3:1 segregation ratio for a single locus. These results indicate that the parents were homozygous for the 2078 mutation at one ACCase gene locus but heterozygous for the 2088 mutation at another ACCase homoeolog (that is, BBC\_ \_ \_ in Table 4).

For the selfed plant containing three mutations, 1781 + 2078 + 2088, the progeny plants segregated for each of the three mutations, indicating that the parent plant was heterozygous at all three mutant loci. Using the 1781 mutation as an example, 55 of 71 progeny plants were scored as RS with the remaining plants lacking the 1781 mutation (Table 4). Pairwise two-point logarithm of the



**Figure 1** Genomic (d)CAPS marker analysis of ACCase mutations showing homozygous susceptible (S) and heterozygous resistant (RS) (allelic and homoeologous) genotypes for the (a) 1781, (b) 2078 and (c) 2088 mutations detected in the parent resistant plants. Note: no homozygous resistant genotypes (RR) are evident, most likely due to fixed homoeologous heterozygosity.

**Table 4** dCAPS and CAPS marker segregation of single-plant-derived (self-pollinated) progeny of the R2 population

Mutation(s) identified in parents	Progeny lines analyzed	No. of progeny plants	Marker-identified genotypes in progeny		Inferred genotypes of the parents	Phenotypic response to diclofop <sup>a</sup>	
			RS (mutation present)	S (mutation absent)		Resistant	Susceptible
1781	1	21	21	0	AA_ _ _ _	21	0
	2	18	18	0		18	0
2078	1	24	24	0	BB_ _ _ _	24	0
	2	20	20	0		20	0
2088	1	18	18	0	CC_ _ _ _	18	0
	2	18	18	0		18	0
1781 + 2088	1	24	24 (1781) 24 (2088)	0 0	AACC_ _ _		
2078 + 2088	1	32	32 (2078) 27 (2088)	0 5	BBC_ _ _		
1781 + 2078 + 2088	1	71	55 (1781) 50 (2078) 49 (2088)	16 21 22	A_ B_ C_		

Abbreviations: dCAPS, derived cleaved amplified polymorphic sequence; RS, resistant mutant and wild-type bands; S, the susceptible wild-type band. Designations of R or S refer to the analyzed mutation-site nucleotides only. A = 1781, B = 2078 and C = 2088 mutation, respectively.

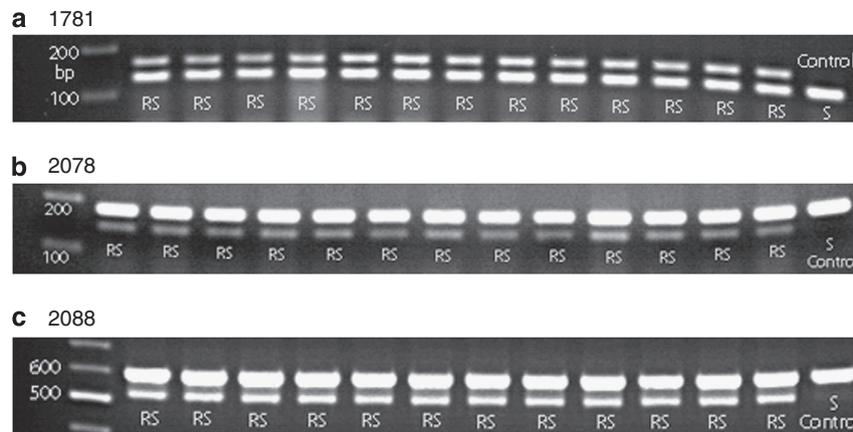
<sup>a</sup>When the genotyping finished, these plants with more than one mutation had already passed the 3- to 4-leaf stage and, therefore, were not treated with diclofop for phenotype assessment.

odds score analyses were performed to detect any genetic linkage among mutations. Maximum logarithm of the odds score values ( $Z_{\max}$ ) were determined for all mutation pairs, and possible linkage was evaluated in both coupling and repulsion phases (with mutations as dominant markers). Results were  $Z_{\max}=0.16$  for 1781 and 2078 mutations linked in repulsion phase, 0.07 for 2078 and 2088 mutations linked in coupling phase and 0.03 for 1781 and 2088 mutations linked in coupling phase (data not shown). Recombination frequencies associated with each  $Z_{\max}$  were 0.42, 0.45 and 0.47, respectively, and these values are near the 0.5 recombination frequency expected of unlinked loci. All  $Z_{\max}$  values were also well below the threshold of 3.0 associated with genetic linkage (Strachan and Read, 1999), indicating that genetic linkage among the three homoeologous ACCase gene loci in wild oat is unlikely and that the three mutations in the parent plant assorted independently.

If the parent plants were heterozygous for each mutation, that is, AaBbCc where lower-case letters designate wild-type alleles, then selfing this parent would result in eight genotype categories among

the progeny when genotyped with dominant d(CAPS) markers (listed in Table 5). The observed number of each genotype in the progeny of the self-pollinated 1781 + 2078 + 2088 mutant plant was found to be consistent with the predicted 27:9:9:9:3:3:3:1 Mendelian disomic segregation ratio for three unlinked loci genotyped using dominant markers ( $P=0.66$ ) (Table 5). This observed segregation pattern is consistent with parent plants being heterozygous for each of the three mutations at each of three unlinked homoeologous ACCase gene loci (A\_B\_C\_ in Table 4).

To further support disomic inheritance expected of *Avena* spp. hexaploids (Thomas, 1992) and associated with the 27:9:9:9:3:3:3:1 segregation ratio, progeny data were tested for goodness of fit to a polysomic inheritance model with random segregation of all six ACCase gene copies in hexaploid wild oat. Chromatid segregation (double reduction) was not considered. As the parent plant had all three mutations with no evidence of genetic linkage, and because two segregates did not carry any of the ACCase mutations (Table 5), the most likely parental genotype under a polysomic inheritance model



**Figure 2** gDNA (d)CAPS marker analysis of progeny plants derived from self-pollinated parents containing the (a) 1781, (b) 2078 or (c) 2088 mutation and showing no marker segregation in the progenies.

**Table 5** Exact test for goodness of fit of observed (d)CAPS marker segregation to a 27:9:9:9:3:3:3:1 ratio expected from disomic inheritance of three unlinked gene loci vs a 147:69:69:69:15:15:15:1 ratio expected from polysomic inheritance of six randomly segregating gene loci, assuming that there is no chromatid segregation (no double reduction)

Mutation/allele	Disomic model				Polysomic model			
	Genotype	Progeny plants			Genotype	Progeny plants		
		Observed	Expected	P		Observed	Expected	P
	if A = 1781, B = 2078, C = 2088				if A = 1781, B = 2078, C = 2088			
1781 + 2078 + 2088	A_B_C_	27	29.9		ABC_ _ _	27	26.1	
1781 + 2078	A_B_cc	12	9.9		AB_ _ _ _	12	12.2	
1781 + 2088	A_bbC_	12	9.9		AC_ _ _ _	12	12.2	
2078 + 2088	aaB_C_	8	9.9		BC_ _ _ _	8	12.2	
1781 only	A_bbcc	5	3.3		A_ _ _ _ _	5	2.7	
2078 only	aaB_cc	4	3.3		B_ _ _ _ _	4	2.7	
2088 only	aabbC_	1	3.3		C_ _ _ _ _	1	2.7	
Nil	aabbcc	2	1.1		wwwwww	2	0.2	
Total		71	71	0.66		71	71	0.009

Abbreviation: dCAPS, derived cleaved amplified polymorphic sequence.

Observations were made among progeny plants derived from a self-pollinated parent plant containing 1781, 2078 and 2088 mutations (A, B and C, respectively). In the disomic model, the parent would be heterozygous for each mutation (AaBbCc × AaBbCc). In the polysomic model, the parent would have all three mutations plus three wild-type alleles (ABCwww × ABCwww). Wild-type alleles are designated a, b or c in the disomic model and w in the polysomic model, and P is probability.

would be ABCwww, where w is a wild-type ACCase gene. The genotype of the two segregates not found to have mutations would be wwwwww, derived from fertilization involving www gametes. Under polysomic inheritance, ABCwww is the most likely genotype of the parent plant because it could generate www gametes while still carrying all three mutant ACCase gene copies. With random segregation of all gene copies, self-pollination of the ABCwww parent would result in an expected segregation ratio of 147:69:69:69:15:15:15:1 when progeny are scored with dominant (d)CAPS markers. Observed progeny marker results were not a good fit to this polysomic inheritance model ( $P=0.009$ ) (Table 5). However, the expected number of wwwwww segregants in the polysomic model was less than one (0.2), which may exaggerate the statistical significance of differences between observed and expected genotype frequencies. On the other hand, expecting 0.2 wwwwww segregants is most consistent with observing either nil or one wwwwww segregant, while we observed two (Table 5). Based on the binomial probability distribution (see Mendenhall, 1987), the probability of observing two or more wwwwww segregants among 71 progeny, where the wwwwww frequency is 1/400 (see the expected polysomic segregation ratio above), is only  $P=0.014$ . Observing the two wwwwww segregants is, therefore, improbable in the polysomic inheritance model.

**Reverse transcriptase-PCR (d)CAPS analysis revealed that three different ACCase mutations can all be transcribed in a single plant**

To confirm that individual plants express all three ACCase resistance mutations, leaf material from nine progeny plants (at the 3 to 4-leaf stage) that were each confirmed to carry all three mutations were used for cDNA-based marker analysis. It was found that the genotype patterns for each of the three mutations were similar when determined using either cDNA or gDNA (note that cDNA and gDNA amplicons have the same sizes as there are no introns within the

amplified fragments) (Figure 3), indicating that genes carrying any of the three ACCase mutations are transcribed. As the three mutations in these plants are each harbored in a different homoeolog, the results also indicate transcription of all three homoeologous ACCase gene copies. However, a much weaker R band was observed relative to the strong S band in both the cDNA and gDNA restriction patterns, especially for the 2078 mutation (Figure 3), probably due to relatively low PCR amplification efficiency for this mutant allele.

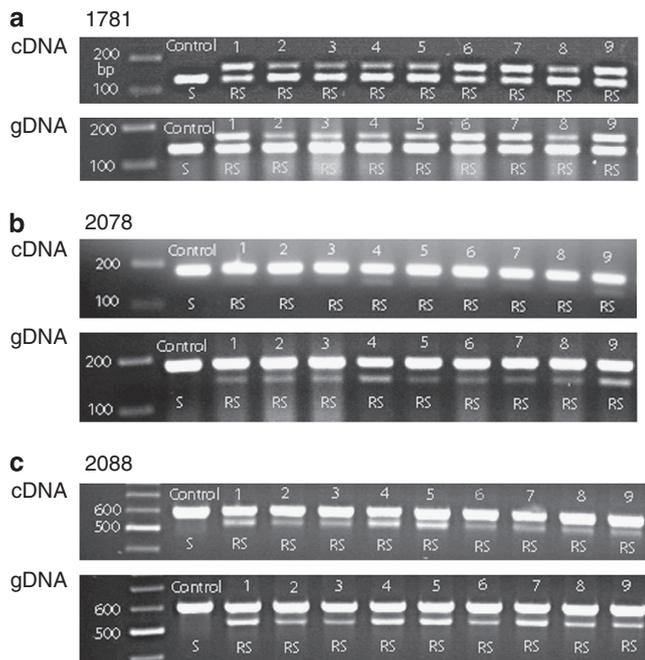
**cDNA cloning further confirmed transcription of the three ACCase mutant alleles in three different *Acc1* gene copies**

Two individual progeny plants that were confirmed to have all three different ACCase resistance mutations in genomic and reverse transcriptase-PCR genotyping (plant 1, 4 in Figure 3) were used for cDNA cloning. The 1122 kb fragment encompassing the 1781, 2078 and 2088 mutation sites was cloned into a plasmid vector and 28 clones with putative inserts from the two plants were picked at random and screened for the 1781, 2078 and 2088 mutations using the three (d)CAPS markers. Thirteen clones had the 1781 mutation, seven had the 2088 mutation, one had the 2078 mutation and seven were wild type. Among the 21 clones that contained mutations, each individual clone was found to have only one of the three mutations (Supplementary Figure S4). Sequencing of the 1781, 2078 and 2088 individual clones further confirmed the three mutations, and alignment of polymorphic sequence regions with the three known *Acc1* homoeologous sequences revealed three resistant alleles (haplotypes). The 2078 haplotype sequence is clearly homologous to *Acc1;3* and the 1781 and 2088 haplotypes are similar to *Acc1;2/Acc1;1* (Figure 4). Therefore, it is clear from molecular marker segregation, reverse transcriptase-PCR marker analysis and cDNA cloning that hexaploid wild oat has three ACCase gene copies, each of which can harbor resistance mutation(s). Importantly, all three copies are expressed as mRNA.

**Assay of *in vitro* ACCase activity revealed that the 1781, 2078 or 2088 mutation in wild oat confers lower-level resistance to ACCase herbicides than in diploid grass species**

To determine if the 1781, 2078 or 2088 resistance mutations individually express as resistant ACCase in wild-oat populations, ACCase activity in the presence or absence of ACCase herbicides was assayed in partially-purified leaf ACCase extracts from susceptible versus resistant plants homozygous, respectively, for each of the three resistance mutations (progeny derived from those in Table 4). The ACCase  $I_{50}$  was estimated for each herbicide and each ACCase resistance mutation (R) versus wild type (S), to give a R/S ratio.

As expected, ACCase from the susceptible plants was confirmed to be sensitive to ACCase herbicides, with  $I_{50}$  values (Table 6) in the range of reported plastidic ACCase values for susceptible grass weed species (Délye, 2005). In contrast, ACCase from resistant plants was significantly less sensitive to ACCase herbicides than the susceptible plants. This reduced sensitivity to ACCase herbicides further confirmed expression of ACCase resistance mutation(s) at the protein level. Importantly, the level of resistance conferred by each mutation as indicated by R/S ratios (1.7–10.8 fold), was generally low (Table 6). Two additional herbicide-susceptible wild-oat populations (*A. fatua* and *A. sterilis*) were used to assess population variation in ACCase herbicide sensitivity, and the diclofop  $I_{50}$  values determined for these two susceptible populations (6.38–6.46  $\mu\text{M}$ , data not included in Table 6) were found similar to that of the susceptible plants originally used. In addition, ACCase isolated from resistant plants (populations R1 and R4) without known ACCase resistance mutations was found



**Figure 3** (d)CAPS marker analysis (a, 1781; b, 2078; c, 2088) of progeny plants carrying three ACCase mutations (1781 + 2078 + 2088) using cDNA versus gDNA and showing the same genotype pattern for each of the mutant alleles.

```

Acc1;1-1      GGAGAACATACATGGAAGTGTGCTATTGCCAGTGCTTATTCTAGGGCATAAGAGGAGAC
2088          GGAGAACATACATGGAAGTGTGCTATTGCCAGTGCTTATTCTAGGGCATAAGAGGAGAC
Acc1;2       GGAGAACATACATGGAAGTGTGCTATTGCCAGTGCTTATTCTAGGGCATAAGAGGAGAC
1781         GGAGAACTTACATGGAAGTGTGCTATTGCCAGTGCTTATTCTAGGGCATAAGAGGAGAC
Acc1;1-2     GGAGAACTTACATGGAAGTGTGCTATTGCCAGTGCTTATTCTAGGGCATAAGAGGAGAC
Acc1;3       GGAGAACATACATGGAAGTGTGCTATTGCCAGTGCTTATTCTAGGGCATAAGAGGAGAC
2078         GGAGAACATACATGGAAGTGTGCTATTGCCAGTGCTTATTCTAGGGCATAAGAGGAGAC
***** * *****

Acc1;1-1      ATTTACTTACATTTGTGAGTGGTCGAACGGTTGGAATAGGAGCCTATCTTGCTCGGCT
2088          ATTTACTTACATTTGTGAGTGGTCGAACGGTTGGAATAGGAGCCTATCTTGCTCGGCT
Acc1;2       ATTTACTTACATTTGTGAGTGGTCGAACGGTTGGAATAGGAGCCTATCTTGCTCGGCT
1781         ATTTACTTACATTTGTGAGTGGTCGAACGGTTGGAATAGGAGCCTATCTTGCTCGGCT
Acc1;1-2     ATTTACTTACATTTGTGAGTGGTCGAACGGTTGGAATAGGAGCCTATCTTGCTCGGCT
Acc1;3       ATTTACTTACATTTGTGAGTGGTCGAACGGTTGGAATAGGAGCCTATCTTGCTCGGCT
2078         ATTTACTTACATTTGTGAGTGGTCGAACGGTTGGAATAGGAGCCTATCTTGCTCGGCT
***** * *****

Acc1;1-1      TGGCATACGGTGCATACAGCGTATTGACCAGCCATTATCTTAACTGGGTTTTCTGCCT
2088          TGGCATACGGTGCATACAGCGTATTGACCAGCCATTATCTTAACTGGGTTTTCTGCCT
Acc1;2       TGGCATACGGTGCATACAGCGTATTGACCAGCCATTATCTTAACTGGGTTTTCTGCCT
1781         TGGCATACGGTGCATACAGCGTATTGACCAGCCATTATCTTAACTGGGTTTTCTGCCT
Acc1;1-2     TGGCATACGGTGCATACAGCGTATTGACCAGCCATTATCTTAACTGGGTTTTCTGCCT
Acc1;3       TGGCATACGGTGCATACAGCGTATTGACCAGCCATTATCTTAACTGGGTTTTCTGCCT
2078         TGGCATACGGTGCATACAGCGTATTGACCAGCCATTATCTTAACTGGGTTTTCTGCCT
***** * *****

Acc1;1-1      GAACAAGCTTCTTGGCCGGGAAGTGTACAGCTCCCACATGCAGTTGGGTGGTCCCAAAT
2088          GAACAAGCTTCTTGGCCGGGAAGTGTACAGCTCCCACATGCAGTTGGGTGGTCCCAAAT
Acc1;2       GAACAAGCTTCTTGGCCGGGAAGTGTACAGCTCCCACATGCAGTTGGGTGGTCCCAAAT
1781         GAACAAGCTTCTTGGCCGGGAAGTGTACAGCTCCCACATGCAGTTGGGTGGTCCCAAAT
Acc1;1-2     GAACAAGCTTCTTGGCCGGGAAGTGTACAGCTCCCACATGCAGTTGGGTGGTCCCAAAT
Acc1;3       GAACAAGCTTCTTGGCCGGGAAGTGTACAGCTCCCACATGCAGTTGGGTGGTCCCAAAT
2078         GAACAAGCTTCTTGGCCGGGAAGTGTACAGCTCCCACATGCAGTTGGGTGGTCCCAAAT
***** * *****

Acc1;1-1      CATGGCTACAAATGGTGTGTCCATCTGACTGTTC
2088          CATGGCTACAAATGGTGTGTCCATCTGACTGTTC
Acc1;2       CATGGCTACAAATGGTGTGTCCATCTGACTGTTC
1781         CATGGCTACAAATGGTGTGTCCATCTGACTGTTC
Acc1;1-2     CATGGCTACAAATGGTGTGTCCATCTGACTGTTC
Acc1;3       CATGGCTACAAATGGTGTGTCCATCTGACTGTTC
2078         CATGGCTACAAATGGTGTGTCCATCTGACTGTTC
***** * *****

```

**Figure 4** Partial sequence alignment (from cloned fragments) showing the three haplotype sequences of the 2088, 1781 and 2078 mutations, corresponding, respectively, to the ACCase gene copies *Acc1;1*, *Acc1;2* and *Acc1;3* in wild oat (Christoffers *et al.*, 2002). Note that the sequence region containing the 1781, 2078 or 2088 codon was not included due to infrequent informative single-nucleotide polymorphisms (SNPs). Boxed regions indicate SNPs discriminating the three haplotypes (GenBank accession AF231335 for *Acc1;1-1*, AF231334 for *Acc1;1-2*, AF231336 for *Acc1;2* and AF464875 for *Acc1;3*).

to be sensitive to fenoxaprop and sethoxydim herbicides, with  $I_{50}$  values of 1.8 and 2.2  $\mu\text{M}$ , respectively (data not included in Table 6).

Reported ACCase  $I_{50}$  R/S ratios for purified resistant diploid *L. rigidum* (Yu *et al.*, 2007) and resistant diploid *Alopecurus myosuroides* (Délye *et al.*, 2005) for the four herbicides (where available) are also cited in Table 6 for comparison. It is evident that the 1781, 2078 or 2088 mutation homozygous in diploid resistant *L. rigidum* or *A. myosuroides* results in much higher ACCase  $I_{50}$  R/S ratios than those displayed by the same mutation(s) in hexaploid wild oat; especially for the 2078 or 2088 mutation.

In order to assess if multiple ACCase gene mutations endow ACCase with higher resistance than single mutations, wild-oat plants having two or more mutations (see Table 4) were assessed for their ACCase sensitivity to diclofop in a separate experiment (experiment II) and the results were compared with the data for the single mutations from experiment I. In general, the ACCase  $I_{50}$  R/S ratios were found to be higher for multiple mutations relative to

single mutations (Table 7). It is noteworthy that the 1781 + 2088 mutants had an  $I_{50}$  R/S ratio of 3.6  $\mu\text{M}$ , which equals the sum of the individual R/S values of the 1781 (2.0  $\mu\text{M}$ ) and 2088 (1.7  $\mu\text{M}$ ) mutants. This indicates an additive resistance interaction among mutant alleles across genomes. Clearly, as will be discussed, the 1781, 2078 or 2088 ACCase mutation alone in hexaploid wild oat does not confer high-level *in vitro* or *in vivo* resistance to ACCase herbicides. It is also notable that in the absence of herbicides, the specific activity of ACCase was much lower for the 2078 or 2088 mutation compared with that of susceptible plants, while the activity of ACCase was unaffected by the 1781 mutation (Table 7). Reduced ACCase activity was also recorded from plants containing two or three resistance mutations (Table 7).

In order to determine if herbicide treatment can induce higher ACCase activity and  $I_{50}$  values, the 2078 or 2088 mutants that were foliar-sprayed with 2000  $\text{g ha}^{-1}$  diclofop-methyl were used for determination of ACCase activity and diclofop  $I_{50}$  values. The results (not

**Table 6** ACCCase  $I_{50}$  values and R/S ratios determined for wild oat (*A. fatua*) homozygous progeny lines (see Table 4) from this study in comparison with the R/S ratios reported for the diploid grass species *Lolium rigidum* and *A. myosuroides* using purified lines that are homozygous for the specific ACCCase mutations

Mutation/mutant allele	Diclofop acid		Fenoxaprop acid		Sethoxydim		Clethodim	
	$I_{50}$ ( $\mu\text{M}$ )	R/S ratio						
<i>Avena</i> spp (hexaploid)								
1781-Leu	16.05 a	2.0	9.79 a	5.3	18.64 a	10.8	2.08 a	3.9
2078-Gly	15.98 a	2.0	7.24 b	3.9	9.79 b	5.7	1.02 b	1.9
2088-Arg	13.54 a	1.7	8.94 ab	4.8	6.10 c	3.5	1.02 b	1.9
Susceptible	7.96 b		1.86 c		1.73 d		0.53 c	
<i>Lolium rigidum</i> <sup>a</sup> (diploid)								
1781-Leu		6				> 100		18
2078-Gly		32				> 100		41
2088-Arg		38				37		48
<i>A. myosuroides</i> <sup>b</sup> (diploid)								
2078-Arg		23		53				36

Abbreviations: ACCCase, acetyl-coenzyme A carboxylase; R/S ratios, resistant/susceptible ratios.

Means with different letters in a column are significantly different ( $\alpha=0.05$ ).

<sup>a</sup>The diclofop and clethodim  $I_{50}$  R/S data are from Yu *et al.* (2007) and the  $I_{50}$  values are not included here because the absolute  $I_{50}$  values of R and S plants can vary because of experimental and assay conditions, while the  $I_{50}$  R/S ratios are more relevant and comparable across species and laboratories. The sethoxydim data is from the current study.

<sup>b</sup>The data are from Délye *et al.* (2005) and the  $I_{50}$  values not included here.

**Table 7** ACCCase activity (in the absence of diclofop) and diclofop  $I_{50}$  values determined for plants containing different resistance mutations/mutant alleles

Mutation/mutant allele	$I_{50}$ values	$I_{50}$ R/S ratio	ACCCase activity (nmol $\text{HCO}_3^- \text{mg}^{-1}$ per protein $\text{min}^{-1}$ )	Activity R/S ratio
<i>Experiment I</i>				
1781-Leu	16.05 a	2.0	1.58 a	0.97
2078-Gly	15.98 a	2.0	0.50 b	0.31
2088-Arg	13.54 a	1.7	0.39 b	0.24
Susceptible	7.96 b		1.63 a	
<i>Experiment II</i>				
1781 + 2078 + 2088	15.92 c	2.6	2.23 b	0.72
2078 + 2088	18.08 b	3.0	1.39 c	0.45
1781 + 2088	21.68 a	3.6	1.34 c	0.43
Susceptible	6.06 d		3.10 a	

Abbreviations: ACCCase, acetyl-coenzyme A carboxylase; R/S ratios, resistant/susceptible ratios. Experiment I was conducted using plant materials at the 3- to 4-leaf stage and experiment II at the 4- to 5-leaf stage. Means with different letters in a column within an experiment are significantly different ( $\alpha=0.05$ ).

shown) were similar to those without diclofop pre-treatment, indicating that herbicide treatment did not induce higher ACCCase activity.

## DISCUSSION

### Multiple ACCCase resistance mutations in individual hexaploid wild-oat plants

Resistance mutation diversity within weed populations and individual plants is important in optimizing weed management strategies. One of the most important findings of this study with hexaploid ACCCase herbicide-resistant wild oat is that individuals can possess multiple

ACCCase resistance mutations among homoeologous genes. While it is common for individuals of cross-pollinated diploid weed species such as *A. myosuroides* and *L. rigidum* (Délye *et al.*, 2005; Yu *et al.*, 2007) to have more than one different ACCCase resistance mutation (compound heterozygosity), multiple mutations have not been reported, to our knowledge, in any field-evolved herbicide-resistant polyploid weed species. Indeed, we found that individuals with two distinct ACCCase mutations are not uncommon (12 of 157 analyzed plants), and we found one individual with three mutations (1781 + 2078 + 2088) (Table 3). The accumulation of three resistance mutations in one plant could occur through independent mutations in all three homoeologous ACCCase gene copies, or due to cross-pollination between individuals carrying different homoeologous ACCCase mutations. Although wild oat is known as a preferentially self-pollinated species, outcrossing can range from 0 to 12% (Imam and Allard, 1965; Murray *et al.*, 2002). Multiple herbicide target-site mutations in individuals of self-pollinated polyploid weed species have been rare thus far, and hence these plants are useful material for genetic and genomic studies for gene expression in polyploid species.

### Genetics and expression of homoeologous ACCCase gene loci in hexaploid wild oat

Hexaploid wild oat has a genome structure of AACCCD, and genome C is clearly distinguishable from A and D (Yang *et al.*, 1999). A previous study of ACCCase herbicide-resistant *A. fatua* identified three homoeologous plastidic ACCCase gene copies, *Acc1;1*, *Acc1;2* and *Acc1;3*, with the 1781 mutation in *Acc1;1* (Christoffers *et al.*, 2002). In comparison with *Avena* spp. diploids, *Acc1;3* was found to likely represent the C genome, while *Acc1;1* and *Acc1;2* likely represent the similar A/D genomes (MJ Christoffers, unpublished data). Subsequent studies with other resistant *A. fatua* populations also found the 2078 or 2088 mutations in *Acc1;1* (Christoffers and Peterson, 2007). However, in studies with resistant *A. sterilis* and *A. fatua* populations, Liu *et al.* (2007) and Beckie *et al.* (2012) indicated that any of the three *Acc1* homoeologs can host ACCCase resistance mutations. In our

study, we identified an individual possessing three distinct ACCase resistance mutations and this provided a unique opportunity to investigate the inheritance of multiple resistance mutations in a polyploid species. We were able to show that individual wild-oat plants can be heterozygous for three different ACCase mutations in each of the three *Acc1* homoeologs, with the 2078 mutation clearly residing in *Acc1;3* (tentatively assigned to the C genome), and the 1781 and 2088 mutations in *Acc1;1/Acc1;2* (likely the A and D genomes) in individuals of the R2 population (Figure 4).

Secondly, our results were consistent with the three homoeologous *Acc1* loci segregating and assorting independently of each other, following classical disomic Mendelian inheritance due to restricted homoeologous pairing and recombination in meiosis (Table 5). Evidence against our data fitting an alternative polysomic inheritance model was highly dependent on the identification of two wild-type plants among segregating progeny. As we cannot be absolutely certain that a genotyping error was not made, we strictly cannot rule out polysomic inheritance based on our data alone. However, the disomic inheritance model favored by the exact test analysis (Table 5) is in line with the established mode of inheritance for hexaploid *Avena* spp. (Thomas, 1992). This disomic segregation pattern in hexaploid *Avena* observed in the current study is also consistent with previous reports on inheritance of ACCase resistance traits (Murray *et al.*, 1995, 1996) and the segregation of wild-type *Acc1;1-1* and resistant *Acc1;1-2* alleles in *A. fatua* (Christoffers *et al.*, 2002).

Thirdly, all three mutant alleles at the three *Acc1* loci are expressed as mRNA (Figure 3, Supplementary Figure S4). Consistent with this conclusion is the recent addition to the GenBank database of 3'-end cDNA sequences for the *Acc1;2* and *Acc1;3* sequences of well-studied UM1 wild oat (HQ244401 and HQ244402, respectively), indicating the transcriptional activity of these genes in another wild-oat population.

It is currently unclear if the three homoeologous ACCase genes in *A. fatua* contribute equally to the transcript pool. In allopolyploid cotton, wheat and other plant species, numerous genes have shown strong expression bias toward one copy (reviewed by Wendel, 2000; Adams, 2007). Many weed species are polyploid and little is currently known about gene expression/silencing or epigenetic regulation in polyploid weeds. In the polyploid *Schoenoplectus mucronatus*, acetolactate synthase gene methylation was detected and epigenetic regulation implicated (Scarabel *et al.*, 2009). Clearly, more research is needed to understand selection and evolution of herbicide resistance genes in polyploid species.

Fixed heterozygosity has also been observed in other polyploid weed species (for example, in *Salsola tragus*, Warwick *et al.*, 2010) in regard to acetolactate synthase resistance mutations. Therefore, PCR-based gene-specific (but not copy-specific) co-dominant molecular markers or sequencing primers designed against conserved regions of herbicide target genes cannot discriminate true (allelic) heterozygotes from homoeoallelic heterozygotes in polyploid species like wild oat. Likewise, detection of true allelic heterozygosity may be similarly complicated in diploid species having multiple herbicide target gene copies. Fixed polymorphism detected by non-copy-specific markers can be indicative of polyploidy or multiple single-genome copies of the gene.

#### ACCase resistance mutations and resistance evolution in hexaploid *Avena*

Most resistance-endowing ACCase gene mutations have been characterized in diploid species (especially *A. myosuroides* and *L. rigidum*) by *in vitro* ACCase assay and whole-plant bioassay using purified lines

homozygous for each of the mutations. Among them, the Ile-1781-Leu mutation was found to confer high-level resistance to the cyclohexanediones herbicide sethoxydim (Table 6) and moderate-level resistance to the aryloxyphenoxypropionates herbicide diclofop (Yu *et al.*, 2007). The Asp-2078-Gly or Cys-2088-Arg mutations conferred high-level resistance to both aryloxyphenoxypropionates and cyclohexanediones herbicides (Délye *et al.*, 2005; Yu *et al.*, 2007; and Table 6). In contrast, *in vitro* inhibition of resistant wild-oat ACCase revealed that each of the mutations conferred much lower-level resistance to the selected ACCase herbicides (Table 6). A similar low-level (2- to 5-fold) resistance to ACCase herbicides was also recently reported for the same 2078 and the Ile-2041-Asn mutation in other *A. fatua* populations (Cruz-Hipoloto *et al.*, 2011). The low resistance observed at the ACCase level for the 1781, 2078 or 2088 resistance mutation in hexaploid wild oat, compared with diploid grass weed species *A. myosuroides* and *L. rigidum* (Table 6), is likely due to polyploidy and dilution of resistant ACCase by susceptible ACCase in wild oat. Assuming equal expression of the three plastidic homoeologous ACCase genes, wild-oat plants homozygous for a given mutation at one ACCase locus would still have two-thirds of the total expressed ACCase as wild-type herbicide sensitive. Furthermore, because ACCase requires two monomers to form a homodimer to yield active ACCase, fully resistant homodimers would likely be further diluted in hexaploid species. Although not examined in this study, differential expression of the three homoeologous ACCase copies is possible and may influence the relative amount of resistant ACCase. If the expression of the ACCase gene copy harboring the ACCase mutation was low compared with other copies, the level of resistance conferred by the mutation would be even further suppressed in wild oat. In contrast, in a diploid homozygous for a given ACCase resistance mutation, all expressed ACCase would be the herbicide resistant isoform. Therefore, an ACCase mutation that confers resistance in diploids may not necessarily endow enough resistance in hexaploids to be selected for in the field (depending on the dominance of the resistance allele, herbicides and herbicide rates). For example, diploid resistant *L. rigidum* survived the Australian field rate of clethodim (60 g ha<sup>-1</sup>) or pinoxaden (30 g ha<sup>-1</sup>) due to the 2078 or 2088 resistance mutation (Yu *et al.*, 2007), but hexaploid wild oat possessing the same mutation rarely survived the herbicide at the same or lower rates (Ahmad-Hamdani *et al.*, 2011; AV Varanasi and MJ Christoffers, submitted).

Whereas gene redundancy in hexaploids may minimize deleterious effects of gene mutations, it may also buffer the effect of resistance mutations (both at target and non-target sites) in herbicide resistance evolution. The dilution effect of resistance mutations by additional susceptible (wild type) homoeologous gene copies may explain in part why wild oat, in comparison with other diploids, is relatively slow to evolve levels of resistance that facilitate identification of resistant populations. In Australia, diclofop was released for *Lolium* and wild oat control in 1978 and despite the presence of both species in many treated fields, resistant *L. rigidum* was reported in 1982 (Heap and Knight, 1982), whereas resistance in wild oat was not reported until 1992 (Mansooji *et al.*, 1992). Very intensive surveys revealed much greater frequency of herbicide resistance in *L. rigidum* than in wild oat in the same region (Owen *et al.*, 2007; Owen and Powles, 2009). Of course, slower resistance evolution in wild oat relative to *L. rigidum* is influenced by several other factors including smaller population sizes, longer seed bank turnover time and extended seed germination (Owen and Powles, 2009). Although self-pollination in wild oat may increase selection for resistance by promoting allelic homozygosity, it may also slow the spread of resistance alleles. Nonetheless, molecular

and genetic evidence from the current study indicates that herbicide resistance and evolution in hexaploid weed species can be considerably more complex than in diploid species.

Although single ACCase mutations in hexaploid wild oat provide relatively low-level resistance to the ACCase herbicides, resistance levels can be significantly increased if individual plants can accumulate multiple ACCase mutations. Based on diclofop  $I_{50}$  R/S ratios, individuals with two mutations had higher resistance than those with a single mutation (Table 7). However, three mutations did not have the expected higher resistance profile, probably because the progeny plants that were used for ACCase assay were mostly heterozygous at two to three *Acc1* loci, with homozygotes for the three mutations at three loci (AABBCC genotype, Table 5) being extremely rare. Although the number of resistant alleles in each case cannot be accurately determined, especially when involving three mutations, the data clearly indicates an additive effect on resistance level across gene copies/genomes, especially when the individuals are homozygous for different mutations. Indeed, the additive effect of resistant acetolactate synthase genes from different genomes has been observed at the acetolactate synthase and whole-plant levels in a mutagenized imidazolinone-resistant wheat line (TealMI15A) (Newhouse *et al.*, 1992; Pozniak and Hucl, 2004; Anderson *et al.*, 2004; Pozniak *et al.*, 2004).

#### Effect of ACCase gene mutations on ACCase activity

Importantly, resistant plants possessing the 2078 or 2088 mutation had <40% of the extractable ACCase activity of susceptible plants (Table 7). Multiple mutations involving 2078 and/or 2088 also had reduced ACCase activity (40–70% of the S, Table 7). This is intriguing given that mutant ACCase isoforms likely account for only about 1/3 of the total ACCase isoforms in the assay mixture. Reduced ACCase activity of the 2078 or 2088 mutation has also been observed in diploid *L. rigidum* and *A. myosuroides* (Yu *et al.*, 2007; Délye *et al.*, 2005). While the single 2078 or 2088 mutation results in significant reduction of *in vitro* ACCase activity in both hexaploid wild oat and diploid *L. rigidum* (Yu *et al.*, 2007), it is unknown if the mutations in wild oat have any fitness cost as they do in *Lolium* (MV Auib, Q Yu, S Powles, unpublished). In *A. myosuroides*, only plants homozygous for the 2078 mutation were found to have a fitness penalty (Menchari *et al.*, 2008). As a polyploid with additional gene copies, wild oat may more readily tolerate mutations that decrease ACCase expression and/or ACCase activity.

#### Non-target-site resistance mechanisms are also evident in resistant wild-oat populations

For fenoxaprop, sethoxydim and clethodim, the ACCase  $I_{50}$  R/S ratios observed for the 1781, 2078 or 2088 mutation at the ACCase level (Table 6) were generally correlated with the LD<sub>50</sub> R/S ratios observed at the whole-plant level (Ahmad-Hamdani *et al.*, 2011). Thus, it's likely that resistance to these three herbicides in these wild-oat populations is largely due to the 1781, 2078 or 2088 mutation. In contrast, high-level resistance to diclofop at the whole plant level (LD<sub>50</sub> R/S ratios >20-fold) (Ahmad-Hamdani *et al.*, 2011), was not consistent with low diclofop ACCase  $I_{50}$  R/S ratios (Table 6), suggesting the possibility of additional non-target-site resistance mechanisms. Some resistant individuals had no known ACCase mutations (Table 3) and their ACCase was herbicide sensitive (see Results). Our recent work on these resistant plants without ACCase resistance mutations demonstrated enhanced herbicide metabolism as a non-target-site resistance mechanism (Ahmad-Hamdani *et al.*, 2012). Both a resistant ACCase and enhanced

rates of herbicide metabolism have been demonstrated in a resistant *A. sterilis* population (Maneechote *et al.*, 1997). Thus, the high-level resistance to some ACCase herbicides (for example, diclofop or fenoxaprop) observed at the population level (Ahmad-Hamdani *et al.*, 2011) is likely due to an ACCase resistance mutation plus metabolic resistance (Ahmad-Hamdani *et al.*, 2012).

In summary, identifying and understanding herbicide resistance and its evolution in hexaploid plant species is more challenging than in diploids. The role of resistance-endowing gene mutations in hexaploid *Avena* species may not be simply inferred from other unrelated diploid species. More complex factors, such as dilution or gene dosage effects and intergenomic gene expression, must be considered in understanding herbicide resistance and evolution in polyploid plant species.

#### DATA ARCHIVING

Partial ACCase sequence alignment showing the three haplotype sequences of the 2088, 1781 and 2078 mutants (Figure 4), and (d)CAPS marker analysis of ACCase resistance mutations in *E. coli* clones (Supplementary Figure S4) have been deposited at Dryad: doi:10.5061/dryad.04562.

#### CONFLICT OF INTEREST

The authors declare no conflict of interest.

#### ACKNOWLEDGEMENTS

This work was supported by the Grains Research and Development Corporation of Australia (grants to the Australian Herbicide Resistance Initiative) and by a PhD scholarship to MS Ahmad-Hamdani from the Ministry of Higher Education Malaysia. We are grateful to Drs Henri Darmency, Christophe Délye and Danica Goggin for valuable comments on the manuscript.

- 
- Adams KL (2007). Evolution of duplicate gene expression in polyploid and hybrid plants. *J Heredity* **98**: 136–141.
- Ahmad-Hamdani MS, Owen MJ, Yu Q, Powles SB (2011). ACCase-inhibiting herbicide resistant wild oat (*Avena fatua* L.) populations from the Western Australian grain belt. *Weed Technol* **26**: 130–136.
- Ahmad-Hamdani MS, Yu Q, Han H, Cawthray GR, Wang SF, Powles SB (2012). Herbicide resistance endowed by enhanced rates of herbicide metabolism in wild oat (*Avena* spp.). *Weed Sci* (in press; doi:10.1614/WS-D-12-00078.1).
- Anderson JA, Matthiesen L, Hegstad J (2004). Resistance to an imidazolinone herbicide is conferred by a gene on chromosome 6DL in the wheat line cv 9804. *Weed Sci* **52**: 83–90.
- Beckie HJ, Warwick SI, Sauder CA (2012). Basis for herbicide resistance in Canadian populations of wild oat (*Avena fatua*). *Weed Sci* **60**: 10–18.
- Bradford MM (1976). A rapid and sensitive method for quantitation of microgram quantities of protein utilising the principle of protein binding. *Anal Biochem* **72**: 248–254.
- Chen ZJ (2007). Genetic and epigenetic mechanisms for gene expression and phenotypic variation in plant polyploids. *Annu Rev Plant Biol* **58**: 377–406.
- Christoffers MJ, Berg ML, Messersmith CG (2002). An isoleucine to leucine mutation in acetyl-CoA carboxylase confers herbicide resistance in wild oat. *Genome* **45**: 1049–1056.
- Christoffers MJ, Pederson SN (2007). Response of wild oat (*Avena fatua*) acetyl-CoA carboxylase mutant to pinoxaden. *Abstr Weed Sci Soc Am* **47**: 258.
- Cocker KM, Coleman JOD, Blair AM, Clarke JH, Moss SR (2000). Biochemical mechanisms of cross-resistance to aryloxyphenoxypropionate and cyclohexanedione herbicides in populations of *Avena* spp. *Weed Res* **40**: 323–334.
- Collavo A, Panozzo S, Lucchesi G, Scarabel L, Sattin M (2011). Characterization and management of *Phalaris paradoxa* resistant to ACCase-inhibitors. *Crop Prot* **30**: 293–299.
- Cruz-Hipolito H, Osuna MD, Dominguez-Valenzuela JA, Espinoza N, De Prado R (2011). Mechanism of resistance to ACCase-inhibiting herbicides in wild oat (*Avena fatua*) from Latin America. *J Agr Food Chem* **59**: 7261–7267.
- Délye C, Zhang XQ, Michel S, Matejick A, Powles SB (2005). Molecular bases for sensitivity to acetyl-coenzyme A carboxylase inhibitors in black-grass. *Plant Physiol* **137**: 794–806.

- Délye C (2005). Weed resistance to acetyl coenzyme A carboxylase inhibitors: an update. *Weed Sci* **53**: 728–746.
- Devine MD, Shimabukuro RH (1994). Resistance to acetyl coenzyme A carboxylase inhibiting herbicides. In: Powles SB, Holtum JAM (eds). *Herbicide Resistance in Plants: Biology and Biochemistry*. Lewis Publishers: Boca Raton, FL, pp 83–139.
- Heap I, Knight R (1982). A population of ryegrass tolerant to the herbicide diclofop-methyl. *J Aust Inst Agric Sci* **48**: 156–157.
- Heap IM (2011). *International Survey of Herbicide-Resistant Weeds*. Available at: <http://www.weedscience.org>. (Accessed March 2012).
- Imam AG, Allard RW (1965). Population studies in predominantly self-pollinated species. VI. Genetic variability between and within natural populations of wild oats from differing habitats in California. *Genetics* **51**: 49–62.
- Jellen EN, Gill BS, Cox TS (1994). Genomic in situ hybridization Differentiates between A/D- and C-genome chromatin and detects intergenomic translocations in polyploid oat species (genus *Avena*). *Genome* **37**: 613–618.
- Kaundun SS, Windass JD (2006). Derived cleaved amplified polymorphic sequence, a simple method to detect a key point mutation conferring acetyl CoA carboxylase inhibitor herbicide resistance in grass weeds. *Weed Sci* **46**: 34–39.
- Kaundun SS (2010). An aspartate to glycine change in the carboxyl transferase domain of acetyl CoA carboxylase and non-target-site mechanism(s) confer resistance to ACCase inhibitor herbicides in a *Lolium multiflorum* population. *Pest Manag Sci* **66**: 1249–1256.
- Liu WJ, Harrison DK, Chalupska D, Gornicki P, O'Donnell CC, Adkins SW *et al.* (2007). Single-site mutations in the carboxyltransferase domain of plastid acetyl-CoA carboxylase confer resistance to grass-specific herbicides. *Proc Natl Acad Sci USA* **104**: 3627–3632.
- Maneeshote C, Holtum JAM, Preston C, Powles SB (1994). Resistant acetyl-CoA carboxylase is a mechanism of herbicide resistance in a biotype of *Avena sterilis* ssp. *ludoviciana*. *Plant Cell Physiol* **35**: 627–635.
- Maneeshote C, Preston C, Powles SB (1997). A diclofop-methyl-resistant *Avena sterilis* biotype with a herbicide-resistance acetyl-coenzyme A carboxylase and enhanced metabolism of diclofop-methyl. *Pestic Sci* **49**: 105–114.
- Mansooji AM, Holtum JA, Boutsalis P, Matthews JM, Powles SB (1992). Resistance to aryloxyphenoxypropionate herbicides in 2 wild oat species (*Avena fatua* and *Avena sterilis* ssp. *ludoviciana*). *Weed Sci* **40**: 599–605.
- Menchari Y, Camilleri C, Michel S, Brunel D, Dessaint F, Le Corre V *et al.* (2006). Weed response to herbicide: regional-scale distribution of herbicide resistance alleles in the grass weed *Alopecurus myosuroides*. *New Phytol* **171**: 861–874.
- Menchari Y, Chauvel B, Darmency H, Délye C (2008). Fitness costs associated with three mutant acetyl-coenzyme A carboxylase alleles endowing herbicide resistance in black-grass *Alopecurus myosuroides*. *J Appl Ecol* **45**: 939–947.
- Mendenhall W (1987). *Introduction to Probability and Statistics*, 7th edn. Duxbury Press: Boston, pp 173–181.
- Murray BG, Brülé-Babel AL, Morrison IN (1996). Two distinct alleles encode for acetyl-CoA carboxylase inhibitor resistance in wild oat (*Avena fatua*). *Weed Sci* **44**: 476–481.
- Murray BG, Morrison IN, Frieses LF (2002). Pollen-mediated gene flow in wild oat. *Weed Sci* **50**: 321–325.
- Murray BG, Morrison IN, Brülé-Babel AL (1995). Inheritance of acetyl-CoA carboxylase inhibitor resistance in wild oat (*Avena fatua*). *Weed Sci* **43**: 233–238.
- Newhouse K, Smith WA, Starrett MA, Schaefer TJ, Singh BK (1992). Tolerance to imidazolinone herbicides in wheat. *Plant Physiol* **100**: 882–886.
- Nikoloudakis N, Katsiotis A (2008). The origin of the C-genome and cytoplasm of *Avena* polyploids. *Theor Appl Genet* **117**: 273–281.
- Nikoloudakis N, Skaracis G, Katsiotis A (2008). Evolutionary insights inferred by molecular analysis of the ITS1-5.8-ITS2 and IGS *Avena* sp. sequences. *Mol Phylogenet Evol* **46**: 102–115.
- Ohsako T, Tominaga T (2007). Nucleotide substitutions in the acetolactate synthase genes of sulfonylurea-resistant biotypes of *Monochoria vaginalis* (Pontederiaceae). *Genes Genet Syst* **82**: 207–215.
- Owen MJ, Powles SB (2009). Distribution and frequency of herbicide-resistant wild oat (*Avena* spp.) across the Western Australian grain belt. *Crop Pasture Sci* **60**: 25–31.
- Powles SB, Yu Q (2010). Evolution in action: plants resistant to herbicides. *Annu Rev Plant Biol* **61**: 317–347.
- Pozniak CJ, Birk IT, O'Donoghue LS, Menard C, Huci PJ, Singh BK (2004). Physiological and molecular characterization of mutation-derived imidazolinone resistance in spring wheat. *Crop Sci* **44**: 1434–1443.
- Pozniak CJ, Huci PJ (2004). Genetic analysis of imidazolinone resistance in mutation-derived lines of common wheat. *Crop Sci* **44**: 23–30.
- Rajhathy T, Morrison JW (1960). Genome homology in the genus *Avena*. *Can J Genet Cytol* **2**: 278–285.
- Scarabel L, Locascio A, Furini A, Sattin M, Varotto S (2009). Characterisation of ALS genes in the polyploid species *Schoenoplectus mucronatus* and implications for resistance management. *Pest Manag Sci* **66**: 337–344.
- Seefeldt SS, Fuerst EP, Gealy DR, Shukala A, Irzyk GP, Devine MD (1996). Mechanisms of resistance to diclofop of two wild oat (*Avena fatua*) biotypes from the Willamette Valley of Oregon. *Weed Sci* **44**: 776–781.
- Scarabel L, Panozzo S, Varotto S, Sattin M (2011). Allelic variation of the ACCase gene and response to ACCase-inhibiting herbicides in pinoxaden-resistant *Lolium* spp. *Pest Manag Sci* **67**: 932–941.
- Shorosh BS, Dixon RA, Ohlrogge JB (1994). Molecular cloning, characterization, and elicitation of acetyl-CoA carboxylase from alfalfa. *Proc Natl Acad Sci USA* **91**: 4323–4327.
- Shukla A, Dupont S, Devine MD (1997). Resistance to ACCase-inhibiting herbicides in wild oat: evidence for target site-based resistance in two biotypes from Canada. *Pestic Biochem Physiol* **57**: 147–155.
- Strachan T, Read AP (1999). *Human Molecular Genetics 2*. Wiley-Liss: New York, pp 274–276.
- Tal A, Rubin B (2004). Molecular characterization and inheritance of resistance to ACCase-inhibiting herbicides in *Lolium rigidum*. *Pest Manag Sci* **60**: 1013–1018.
- Thomas T (1992). Cytogenetics of *Avena*. In: Marshall HG, Sorrells ME (eds). *Oat Science and Technology*. American Society of Agronomy and Crop Science Society of America: Madison, WI, pp 473–507.
- Uchino A, Ogata S, Kohara H, Yoshida S, Yoshioka T, Watanabe H (2007). Molecular basis of diverse responses to acetolactate synthase-inhibiting herbicides in sulfonylurea-resistant biotypes of *Schoenoplectus juncooides*. *Weed Biol Manag* **7**: 89–96.
- Warwick S, Sauder CA, Beckie HJ (2010). Acetolactate synthase (ALS) target-site mutations in ALS inhibitor-resistant Russian thistle (*Salsola tragus*). *Weed Sci* **58**: 244–251.
- Wendel JF (2000). Genome evolution in polyploids. *Plant Mol Biol* **42**: 225–249.
- Yang Q, Hanson L, Bennett MD, Leitch IJ (1999). Genome structure and evolution in the allohexaploid weed *Avena fatua* L. (Poaceae). *Genome* **42**: 512–518.
- Yu Q, Collavo A, Zheng MQ, Owen M, Sattin M, Powles SB (2007). Diversity of acetyl-coenzyme A carboxylase mutations in resistant *Lolium* populations: evaluation using clethodim. *Plant Physiol* **145**: 547–555.
- Yu Q, Friesen S, Zhang XQ, Powles SB (2004). Tolerance to acetolactate synthase and acetyl-coenzyme A carboxylase inhibiting herbicides in *Vulpia bromoides* is conferred by two co-existing resistance mechanisms. *Pestic Biochem Physiol* **21**: 21–30.
- Yu Q, Han HP, Powles SB (2008). Mutations of the ALS gene endowing resistance to ALS-inhibiting herbicides in *Lolium rigidum* populations. *Pest Manag Sci* **64**: 1229–1236.

Supplementary Information accompanies the paper on Heredity website (<http://www.nature.com/hdy>)